



Figure 1. Desorption gas chromatograms from 1-h collections of flow tube gases. Lower trace: no ions present. Upper trace: ions present. All peaks were characterized by their mass spectrum and are identified as (A) NF_3 ; (B) $\text{C}_6\text{H}_5\text{COOH}$; (C) CH_2Cl_2 ; (D) $(\text{CH}_3)_2\text{CO}$; (E) $(\text{CH}_3)_2\text{CHOH}$; (F) CHCl_3 ; (G) $\text{CF}_2\text{ClCCl}_2\text{F}$; (H) $\text{CH}_2=\text{CHCH}_2\text{C}-\text{H}_2\text{Cl}$; (I) CH_3CCl_3 ; (J) CCl_4 ; (K) $\text{CH}_3\text{CHClCH}_2\text{CH}_3$; (L) $(\text{CH}_3)_3\text{CCH}_2\text{Cl}$.

to the flow reactor. By addition of approximately 2 mtorr of NH_3 to the buffer gas, all positive ion reactions of **1** were suppressed.

The sampling port is 20 cm downstream from the neutral addition inlet. A 10-mm id borosilicate glass collection column was packed with glass wool that had been coated with 2,6-diphenyl-*p*-phenylene oxide-polymer (Tenax-GC⁶). This porous polymer is a relatively inert, reversible sorbent for collection of volatile, environmental samples.⁷ Polymer-coated glass wool provides an inert surface for low-temperature condensation of volatile compounds. A small fraction (1.5×10^{-4}) of the gases in the flow tube was diverted to the cooled (-198°C) collection column. A fluoride ion density of $3 \times 10^8 \text{ cm}^{-3}$ produces a like number of **2**, and, with 100% sampling efficiency, we should find about 8 ng of neopentyl fluoride in 1 h.

After collection (15 min to 1 h), the sampling column was removed and connected to the inlet port of a HP 5982A GC/MS/DS. The neutrals were then backflushed at 190°C and focused at -78°C onto the head of a $1/4 \text{ in.} \times 6 \text{ ft}$ chromatographic column (0.1% SP1000 on Carbowax C). Subsequent temperature-programmed elution coupled with full mass scanning detection by the MS allow unambiguous identification of collected species. Comparison with appropriate standards allows quantitative determination of the collected products.⁸

Figure 1 depicts the results of a typical GC/MS study for a 1-h collection. The lower trace shows neutrals collected with the ion source off (all reagents present but no ions). The upper trace shows a sample collected in the presence of F^- ; the only additional peak is due to 6 ng of neopentyl fluoride. A full mass scan of the new peak at 8.3 min absolutely identifies it as the reaction product. Methylene chloride, acetone, and 2-propanol (peaks C, D, and E) are ubiquitous solvents, and all other peaks are due to trace-level impurities in the starting material (**1**). This proves that **2** has been produced by a chemical reaction in our flow tube.

Some serious objections remain. How can we be sure that surface reactions (either on the walls of the flow tube or on the collection column) were not responsible for **2**? Can we say that **2** is produced by a negative ion reaction such as in eq 1 and not an atom or radical reaction? We certainly produce F and NF_2 at the ion source as well as F^- .

Two control experiments answered these questions. Neopentyl chloride alone was injected into the flowing afterglow with no ions

present. After deposition of **1** onto the cooled collection column for 30 min, the neopentyl chloride was turned off; F^- was then generated from NF_3 , and sampling was continued for 1 h. A chromatogram of this sample shows that **2** is not present. Therefore, our neopentyl fluoride was not produced by surface reactions. Our afterglow is equipped with an ion shutter.^{5c} This is a tungsten mesh astride the flow tube. When biased at -100 V , the quadrupole MS indicated that all negative ions were blocked, and yet all neutrals (NF_3 , F, NF_2 , etc.) were free to pass. We then added **1** downstream from the shutter and collected as before. Again, we could not detect the presence of **2** with our GC/MS/DS (sensitivity for **2** around 100 pg). Both F^- and **1** must interact in the flow tube to produce neopentyl fluoride; radical reactions do not produce our product. These experiments establish that eq 1 is correct as written by Olmstead and Brauman.³

Neutral product analysis uniquely characterizes the chemistry of gas-phase ions. This powerful new technique now clears the way for a more complete understanding of ion-molecule reactions in the absence of solvent.

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Facile Syntheses of Tetraalkylchlorin and Tetraalkylporphyrin Complexes and Comparison of the Structures of the Tetramethylchlorin and Tetramethylporphyrin Complexes of Nickel(II)

Sir:

We report facile syntheses of meso-substituted tetraalkylchlorin and tetraalkylporphyrin complexes of some first-row transition metals and provide a comparison of the structures of $\text{Ni}(\text{TMC})$ and $\text{Ni}(\text{TMP})$, where $\text{TMC} = 5,10,15,20$ -tetramethylchlorinato and $\text{TMP} = 5,10,15,20$ -tetramethylporphyrinato. The use of metalloporphyrins as models for coordination sites in metallo-biomolecules is now of recognized importance.¹ That reduced hemes occur in certain biological systems^{2,3} has led to interest in the synthesis of model systems^{4,5} involving reduced porphyrins, e.g., chlorins, bacteriochlorins, and isobacteriochlorins. In no instance has there been available an easy route to analytically and isomerically pure samples of metal complexes of reduced porphyrins. To this end, we have turned our attention to the synthesis of the simple TMP and TMC and related tetraalkyl systems.

All suggested syntheses for $\text{H}_2\text{TMP}^{6-8}$ report problems with "contamination" by H_2TMC , probably because H_2TMC has a higher oxidation potential than tetraphenyl- or octaethylchlorin. We find that a template synthesis increases the "contamination" of TMP by TMC to make it the major product. Reaction 1 with $\text{R} = \text{Me}$ affords different $\text{M}(\text{TMC})/\text{M}(\text{TMP})$ ratios, depending upon the metal M and the amount of anhydride present. In the presence of 2% acetic anhydride, the following products⁹ were

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